

GEORGIAN MEDICAL NEWS

ISSN 1512-0112

No 1 (310) Январь 2021

ТБИЛИСИ - NEW YORK



ЕЖЕМЕСЯЧНЫЙ НАУЧНЫЙ ЖУРНАЛ

Медицинские новости Грузии
საქართველოს სამედიცინო სიახლენი

GEORGIAN MEDICAL NEWS

No 1 (310) 2021

Published in cooperation with and under the patronage
of the Tbilisi State Medical University

Издается в сотрудничестве и под патронажем
Тбилисского государственного медицинского университета

გამოიცემა თბილისის სახელმწიფო სამედიცინო უნივერსიტეტთან
თანამშრომლობითა და მისი პატრონაჟით

**ЕЖЕМЕСЯЧНЫЙ НАУЧНЫЙ ЖУРНАЛ
ТБИЛИСИ - НЬЮ-ЙОРК**

GMN: Georgian Medical News is peer-reviewed, published monthly journal committed to promoting the science and art of medicine and the betterment of public health, published by the GMN Editorial Board and The International Academy of Sciences, Education, Industry and Arts (U.S.A.) since 1994. **GMN** carries original scientific articles on medicine, biology and pharmacy, which are of experimental, theoretical and practical character; publishes original research, reviews, commentaries, editorials, essays, medical news, and correspondence in English and Russian.

GMN is indexed in MEDLINE, SCOPUS, PubMed and VINITI Russian Academy of Sciences. The full text content is available through EBSCO databases.

GMN: Медицинские новости Грузии - ежемесячный рецензируемый научный журнал, издаётся Редакционной коллегией и Международной академией наук, образования, искусств и естествознания (IASEIA) США с 1994 года на русском и английском языках в целях поддержки медицинской науки и улучшения здравоохранения. В журнале публикуются оригинальные научные статьи в области медицины, биологии и фармации, статьи обзорного характера, научные сообщения, новости медицины и здравоохранения.

Журнал индексируется в MEDLINE, отражён в базе данных SCOPUS, PubMed и ВИНТИ РАН. Полнотекстовые статьи журнала доступны через БД EBSCO.

GMN: Georgian Medical News – საქართველოს სამედიცინო სიახლენი – არის ყოველთვიური სამეცნიერო სამედიცინო რეცენზირებადი ჟურნალი, გამოიცემა 1994 წლიდან, წარმოადგენს სარედაქციო კოლეგიისა და აშშ-ის მეცნიერების, განათლების, ინდუსტრიის, ხელოვნებისა და ბუნებისმეტყველების საერთაშორისო აკადემიის ერთობლივ გამოცემას. GMN-ში რუსულ და ინგლისურ ენებზე ქვეყნდება ექსპერიმენტული, თეორიული და პრაქტიკული ხასიათის ორიგინალური სამეცნიერო სტატიები მედიცინის, ბიოლოგიისა და ფარმაციის სფეროში, მიმოხილვითი ხასიათის სტატიები.

ჟურნალი ინდექსირებულია MEDLINE-ის საერთაშორისო სისტემაში, ასახულია SCOPUS-ის, PubMed-ის და ВИНТИ РАН-ის მონაცემთა ბაზებში. სტატიების სრული ტექსტი ხელმისაწვდომია EBSCO-ს მონაცემთა ბაზებშიდან.

МЕДИЦИНСКИЕ НОВОСТИ ГРУЗИИ

Ежемесячный совместный грузино-американский научный электронно-печатный журнал
Агентства медицинской информации Ассоциации деловой прессы Грузии,
Международной академии наук, индустрии, образования и искусств США.
Издается с 1994 г., распространяется в СНГ, ЕС и США

ГЛАВНЫЙ РЕДАКТОР

Николай Пирцхалаишвили

НАУЧНЫЙ РЕДАКТОР

Елене Гиоргадзе

ЗАМЕСТИТЕЛЬ ГЛАВНОГО РЕДАКТОРА

Нино Микаберидзе

НАУЧНО-РЕДАКЦИОННЫЙ СОВЕТ

Зураб Вадачкориа - председатель Научно-редакционного совета

Михаил Бахмутский (США), Александр Геннинг (Германия), Амиран Гамкрелидзе (Грузия),
Константин Кипиани (Грузия), Георгий Камкамидзе (Грузия),
Паата Куртанидзе (Грузия), Вахтанг Масхулия (Грузия),
Тенгиз Ризнис (США), Реваз Сепиашвили (Грузия), Дэвид Элуа (США)

НАУЧНО-РЕДАКЦИОННАЯ КОЛЛЕГИЯ

Константин Кипиани - председатель Научно-редакционной коллегии

Архимандрит Адам - Вахтанг Ахаладзе, Амиран Антадзе, Нелли Антелава, Тенгиз Асатиани,
Гия Берадзе, Рима Бериашвили, Лео Бокерия, Отар Герзмава, Лиана Гогиашвили, Нодар Гогебашвили,
Николай Гонгадзе, Лия Дваладзе, Тамар Долиашвили, Манана Жвания, Тамар Зерекидзе,
Ирина Квачадзе, Нана Квирквелия, Зураб Кеванишвили, Гурам Кикнадзе, Димитрий
Кордзаиа, Теймураз Лежава, Нодар Ломидзе, Джанлуиджи Мелотти, Марина Мамаладзе,
Караман Пагава, Мамука Пирцхалаишвили, Анна Рехвиашвили, Мака Сологашвили, Рамаз Хецуриани,
Рудольф Хохенфеллнер, Кахабер Челидзе, Тинатин Чиковани, Арчил Чхотуа,
Рамаз Шенгелия, Кетеван Эбралидзе

Website:

www.geomednews.org

The International Academy of Sciences, Education, Industry & Arts. P.O.Box 390177,
Mountain View, CA, 94039-0177, USA. Tel/Fax: (650) 967-4733

Версия: печатная. **Цена:** свободная.

Условия подписки: подписка принимается на 6 и 12 месяцев.

По вопросам подписки обращаться по тел.: 293 66 78.

Контактный адрес: Грузия, 0177, Тбилиси, ул. Асатиани 7, IV этаж, комната 408
тел.: 995(32) 254 24 91, 5(55) 75 65 99

Fax: +995(32) 253 70 58, e-mail: ninomikaber@geomednews.com; nikopir@geomednews.com

По вопросам размещения рекламы обращаться по тел.: 5(99) 97 95 93

© 2001. Ассоциация деловой прессы Грузии

© 2001. The International Academy of Sciences,
Education, Industry & Arts (USA)

GEORGIAN MEDICAL NEWS

Monthly Georgia-US joint scientific journal published both in electronic and paper formats of the Agency of Medical Information of the Georgian Association of Business Press; International Academy of Sciences, Education, Industry and Arts (USA).
Published since 1994. Distributed in NIS, EU and USA.

EDITOR IN CHIEF

Nicholas Pirtskhalaishvili

SCIENTIFIC EDITOR

Elene Giorgadze

DEPUTY CHIEF EDITOR

Nino Mikaberidze

SCIENTIFIC EDITORIAL COUNCIL

Zurab Vadachkoria - Head of Editorial council

Michael Bakhmutsky (USA), Alexander Gënning (Germany),
Amiran Gamkrelidze (Georgia), David Elua (USA),
Konstantin Kipiani (Georgia), Giorgi Kamkamidze (Georgia), Paata Kurtanidze (Georgia),
Vakhtang Maskhulia (Georgia), Tengiz Riznis (USA), Revaz Sepiashvili (Georgia)

SCIENTIFIC EDITORIAL BOARD

Konstantin Kipiani - Head of Editorial board

Archimandrite Adam - Vakhtang Akhaladze, Amiran Antadze, Nelly Antelava,
Tengiz Asatiani, Gia Beradze, Rima Beriashvili, Leo Bokeria, Kakhaber Chelidze,
Tinatin Chikovani, Archil Chkhotua, Lia Dvaladze, Tamar Doliashvili, Ketevan Ebralidze,
Otar Gerzmava, Liana Gogiashvili, Nodar Gogebashvili, Nicholas Gongadze,
Rudolf Hohenfellner, Zurab Kevanishvili, Ramaz Khetsuriani, Guram Kiknadze,
Dimitri Kordzaia, Irina Kvachadze, Nana Kvirkvelia, Teymuraz Lezhava, Nodar Lomidze, Marina
Mamaladze, Gianluigi Melotti, Kharaman Pagava, Mamuka Pirtskhalaishvili,
Anna Rekhviashvili, Maka Sologhashvili, Ramaz Shengelia, Tamar Zerekidze, Manana Zhvania

CONTACT ADDRESS IN TBILISI

GMN Editorial Board
7 Asatiani Street, 4th Floor
Tbilisi, Georgia 0177

Phone: 995 (32) 254-24-91
995 (32) 253-70-58
Fax: 995 (32) 253-70-58

CONTACT ADDRESS IN NEW YORK

NINITEX INTERNATIONAL, INC.
3 PINE DRIVE SOUTH
ROSLYN, NY 11576 U.S.A.

Phone: +1 (917) 327-7732

WEBSITE

www.geomednews.org

К СВЕДЕНИЮ АВТОРОВ!

При направлении статьи в редакцию необходимо соблюдать следующие правила:

1. Статья должна быть представлена в двух экземплярах, на русском или английском языках, напечатанная через **полтора интервала на одной стороне стандартного листа с шириной левого поля в три сантиметра**. Используемый компьютерный шрифт для текста на русском и английском языках - **Times New Roman (Кириллица)**, для текста на грузинском языке следует использовать **AcadNusx**. Размер шрифта - **12**. К рукописи, напечатанной на компьютере, должен быть приложен CD со статьей.

2. Размер статьи должен быть не менее десяти и не более двадцати страниц машинописи, включая указатель литературы и резюме на английском, русском и грузинском языках.

3. В статье должны быть освещены актуальность данного материала, методы и результаты исследования и их обсуждение.

При представлении в печать научных экспериментальных работ авторы должны указывать вид и количество экспериментальных животных, применявшиеся методы обезболивания и усыпления (в ходе острых опытов).

4. К статье должны быть приложены краткое (на полстраницы) резюме на английском, русском и грузинском языках (включающее следующие разделы: цель исследования, материал и методы, результаты и заключение) и список ключевых слов (key words).

5. Таблицы необходимо представлять в печатной форме. Фотокопии не принимаются. **Все цифровые, итоговые и процентные данные в таблицах должны соответствовать таковым в тексте статьи**. Таблицы и графики должны быть озаглавлены.

6. Фотографии должны быть контрастными, фотокопии с рентгенограмм - в позитивном изображении. Рисунки, чертежи и диаграммы следует озаглавить, пронумеровать и вставить в соответствующее место текста **в tiff формате**.

В подписях к микрофотографиям следует указывать степень увеличения через окуляр или объектив и метод окраски или импрегнации срезов.

7. Фамилии отечественных авторов приводятся в оригинальной транскрипции.

8. При оформлении и направлении статей в журнал МНГ просим авторов соблюдать правила, изложенные в «Единых требованиях к рукописям, представляемым в биомедицинские журналы», принятых Международным комитетом редакторов медицинских журналов - <http://www.spinesurgery.ru/files/publish.pdf> и http://www.nlm.nih.gov/bsd/uniform_requirements.html В конце каждой оригинальной статьи приводится библиографический список. В список литературы включаются все материалы, на которые имеются ссылки в тексте. Список составляется в алфавитном порядке и нумеруется. Литературный источник приводится на языке оригинала. В списке литературы сначала приводятся работы, написанные знаками грузинского алфавита, затем кириллицей и латиницей. Ссылки на цитируемые работы в тексте статьи даются в квадратных скобках в виде номера, соответствующего номеру данной работы в списке литературы. Большинство цитированных источников должны быть за последние 5-7 лет.

9. Для получения права на публикацию статья должна иметь от руководителя работы или учреждения визу и сопроводительное отношение, написанные или напечатанные на бланке и заверенные подписью и печатью.

10. В конце статьи должны быть подписи всех авторов, полностью приведены их фамилии, имена и отчества, указаны служебный и домашний номера телефонов и адреса или иные координаты. Количество авторов (соавторов) не должно превышать пяти человек.

11. Редакция оставляет за собой право сокращать и исправлять статьи. Корректур авторам не высылаются, вся работа и сверка проводится по авторскому оригиналу.

12. Недопустимо направление в редакцию работ, представленных к печати в иных издательствах или опубликованных в других изданиях.

При нарушении указанных правил статьи не рассматриваются.

REQUIREMENTS

Please note, materials submitted to the Editorial Office Staff are supposed to meet the following requirements:

1. Articles must be provided with a double copy, in English or Russian languages and typed or computer-printed on a single side of standard typing paper, with the left margin of 3 centimeters width, and 1.5 spacing between the lines, typeface - **Times New Roman (Cyrillic)**, print size - 12 (referring to Georgian and Russian materials). With computer-printed texts please enclose a CD carrying the same file titled with Latin symbols.

2. Size of the article, including index and resume in English, Russian and Georgian languages must be at least 10 pages and not exceed the limit of 20 pages of typed or computer-printed text.

3. Submitted material must include a coverage of a topical subject, research methods, results, and review.

Authors of the scientific-research works must indicate the number of experimental biological species drawn in, list the employed methods of anesthetization and soporific means used during acute tests.

4. Articles must have a short (half page) abstract in English, Russian and Georgian (including the following sections: aim of study, material and methods, results and conclusions) and a list of key words.

5. Tables must be presented in an original typed or computer-printed form, instead of a photocopied version. **Numbers, totals, percentile data on the tables must coincide with those in the texts of the articles.** Tables and graphs must be headed.

6. Photographs are required to be contrasted and must be submitted with doubles. Please number each photograph with a pencil on its back, indicate author's name, title of the article (short version), and mark out its top and bottom parts. Drawings must be accurate, drafts and diagrams drawn in Indian ink (or black ink). Photocopies of the X-ray photographs must be presented in a positive image in **tiff format**.

Accurately numbered subtitles for each illustration must be listed on a separate sheet of paper. In the subtitles for the microphotographs please indicate the ocular and objective lens magnification power, method of coloring or impregnation of the microscopic sections (preparations).

7. Please indicate last names, first and middle initials of the native authors, present names and initials of the foreign authors in the transcription of the original language, enclose in parenthesis corresponding number under which the author is listed in the reference materials.

8. Please follow guidance offered to authors by The International Committee of Medical Journal Editors guidance in its Uniform Requirements for Manuscripts Submitted to Biomedical Journals publication available online at: http://www.nlm.nih.gov/bsd/uniform_requirements.html
http://www.icmje.org/urm_full.pdf

In GMN style for each work cited in the text, a bibliographic reference is given, and this is located at the end of the article under the title "References". All references cited in the text must be listed. The list of references should be arranged alphabetically and then numbered. References are numbered in the text [numbers in square brackets] and in the reference list and numbers are repeated throughout the text as needed. The bibliographic description is given in the language of publication (citations in Georgian script are followed by Cyrillic and Latin).

9. To obtain the rights of publication articles must be accompanied by a visa from the project instructor or the establishment, where the work has been performed, and a reference letter, both written or typed on a special signed form, certified by a stamp or a seal.

10. Articles must be signed by all of the authors at the end, and they must be provided with a list of full names, office and home phone numbers and addresses or other non-office locations where the authors could be reached. The number of the authors (co-authors) must not exceed the limit of 5 people.

11. Editorial Staff reserves the rights to cut down in size and correct the articles. Proof-sheets are not sent out to the authors. The entire editorial and collation work is performed according to the author's original text.

12. Sending in the works that have already been assigned to the press by other Editorial Staffs or have been printed by other publishers is not permissible.

**Articles that Fail to Meet the Aforementioned
Requirements are not Assigned to be Reviewed.**

ავტორთა საქურაღებოლ!

რედაქციაში სტატიის წარმოდგენისას საჭიროა დაიცვათ შემდეგი წესები:

1. სტატია უნდა წარმოადგინოთ 2 ცალად, რუსულ ან ინგლისურ ენებზე დაბეჭდილი სტანდარტული ფურცლის 1 გვერდზე, 3 სმ სიგანის მარცხენა ველისა და სტრიქონებს შორის 1,5 ინტერვალის დაცვით. გამოყენებული კომპიუტერული შრიფტი რუსულ და ინგლისურენოვან ტექსტებში - **Times New Roman (Кириллица)**, ხოლო ქართულენოვან ტექსტში საჭიროა გამოვიყენოთ **AcadNusx**. შრიფტის ზომა – 12. სტატიას თან უნდა ახლდეს CD სტატიით.

2. სტატიის მოცულობა არ უნდა შეადგენდეს 10 გვერდზე ნაკლებს და 20 გვერდზე მეტს ლიტერატურის სიის და რეზიუმეების (ინგლისურ, რუსულ და ქართულ ენებზე) ჩათვლით.

3. სტატიაში საჭიროა გაშუქდეს: საკითხის აქტუალობა; კვლევის მიზანი; საკვლევი მასალა და გამოყენებული მეთოდები; მიღებული შედეგები და მათი განსჯა. ექსპერიმენტული ხასიათის სტატიების წარმოდგენისას ავტორებმა უნდა მიუთითონ საექსპერიმენტო ცხოველების სახეობა და რაოდენობა; გაუტკივარებისა და დაძინების მეთოდები (მწვავე ცდების პირობებში).

4. სტატიას თან უნდა ახლდეს რეზიუმე ინგლისურ, რუსულ და ქართულ ენებზე არანაკლებ ნახევარი გვერდის მოცულობისა (სათაურის, ავტორების, დაწესებულების მითითებით და უნდა შეიცავდეს შემდეგ განყოფილებებს: მიზანი, მასალა და მეთოდები, შედეგები და დასკვნები; ტექსტუალური ნაწილი არ უნდა იყოს 15 სტრიქონზე ნაკლები) და საკვანძო სიტყვების ჩამონათვალი (key words).

5. ცხრილები საჭიროა წარმოადგინოთ ნაბეჭდი სახით. ყველა ციფრული, შემაჯამებელი და პროცენტული მონაცემები უნდა შეესაბამებოდეს ტექსტში მოყვანილს.

6. ფოტოსურათები უნდა იყოს კონტრასტული; სურათები, ნახაზები, დიაგრამები - დასათაურებული, დანომრილი და სათანადო ადგილას ჩასმული. რენტგენოგრაფიების ფოტოასლები წარმოადგინეთ პოზიტიური გამოსახულებით **tiff** ფორმატში. მიკროფოტოსურათების წარწერებში საჭიროა მიუთითოთ ოკულარის ან ობიექტივის საშუალებით გადიდების ხარისხი, ანათალების შედეგების ან იმპრეგნაციის მეთოდი და აღნიშნოთ სურათის ზედა და ქვედა ნაწილები.

7. სამამულო ავტორების გვარები სტატიაში აღინიშნება ინიციალების თანდართვით, უცხოურისა – უცხოური ტრანსკრიპციით.

8. სტატიას თან უნდა ახლდეს ავტორის მიერ გამოყენებული სამამულო და უცხოური შრომების ბიბლიოგრაფიული სია (ბოლო 5-8 წლის სიღრმით). ანბანური წყობით წარმოდგენილ ბიბლიოგრაფიულ სიაში მიუთითეთ ჯერ სამამულო, შემდეგ უცხოელი ავტორები (გვარი, ინიციალები, სტატიის სათაური, ჟურნალის დასახელება, გამოცემის ადგილი, წელი, ჟურნალის №, პირველი და ბოლო გვერდები). მონოგრაფიის შემთხვევაში მიუთითეთ გამოცემის წელი, ადგილი და გვერდების საერთო რაოდენობა. ტექსტში კვადრატულ ფხიხლებში უნდა მიუთითოთ ავტორის შესაბამისი N ლიტერატურის სიის მიხედვით. მიზანშეწონილია, რომ ციტირებული წყაროების უმეტესი ნაწილი იყოს 5-6 წლის სიღრმის.

9. სტატიას თან უნდა ახლდეს: ა) დაწესებულების ან სამეცნიერო ხელმძღვანელის წარდგინება, დამოწმებული ხელმოწერითა და ბეჭდით; ბ) დარგის სპეციალისტის დამოწმებული რეცენზია, რომელშიც მითითებული იქნება საკითხის აქტუალობა, მასალის საკმაობა, მეთოდის სანდოობა, შედეგების სამეცნიერო-პრაქტიკული მნიშვნელობა.

10. სტატიის ბოლოს საჭიროა ყველა ავტორის ხელმოწერა, რომელთა რაოდენობა არ უნდა აღემატებოდეს 5-ს.

11. რედაქცია იტოვებს უფლებას შეასწოროს სტატია. ტექსტზე მუშაობა და შეჯერება ხდება საავტორო ორიგინალის მიხედვით.

12. დაუშვებელია რედაქციაში ისეთი სტატიის წარდგენა, რომელიც დასაბეჭდად წარდგენილი იყო სხვა რედაქციაში ან გამოქვეყნებული იყო სხვა გამოცემებში.

აღნიშნული წესების დარღვევის შემთხვევაში სტატიები არ განიხილება.

Содержание:

Taner Demirci, Hasret Cengiz, Sedat Cetin, Ceyhun Varim, Gizem Karatas Kılıçcioğlu MYELOLIPOMA COEXISTENCE WITH GLUCOCORTICOID AND ANDROGEN SECRETING ADRENOCORTICAL CARCINOMA: SLOW AND BENIGN CLINICAL COURSE.....	7
Русин В.И., Русин В.В., Горленко Ф.В., Добош В.М., Лопит М.М. ИЗОЛИРОВАННАЯ ПРОФУНДОПЛАСТИКА (ДИФФЕРЕНЦИРОВАННЫЙ ВЫБОР).....	11
Зубач О.Б., Григорьева Н.В., Поворозник В.В. 10-ЛЕТНЯЯ ЛЕТАЛЬНОСТЬ У ПАЦИЕНТОВ ПОСЛЕ ПЕРЕЛОМОВ ПРОКСИМАЛЬНОГО ОТДЕЛА БЕДРЕННОЙ КОСТИ.....	19
Zenaishvili M., Japaridze Sh., Tushishvili A., Davitashvili O., Kevanishvili Z. STUTTERING: INITIATING FACTORS, EVOLUTION, HEALING PERSPECTIVES.....	23
Hirna H., Kostyshyn I., Rozhko M., Levandovskiy R., Nakashidze G. ANALYSIS OF IMMUNE CHANGES AND THEIR ROLE IN THE DEVELOPMENT OF ORAL AND OROPHARYNGEAL CANCER	29
Tsitadze T., Puturidze S., Lomidze T., Margvelashvili V., Kalandadze M. PREVALENCE AND RISK-FACTORS OF BRUXISM IN CHILDREN AND ADOLESCENT POPULATION AND ITS IMPACT ON QUALITY OF LIFE (REVIEW).....	36
Solovyeva Z., Zaporozhskaya-Abramova E., Adamchik A., Gushchin A., Risovanniy S., Manukyan I. COMPARATIVE EVALUATION OF THE CLINICAL EFFICACY OF MODERN REMINERALIZING DRUGS IN THE TREATMENT OF ENAMEL CARIES (FOCAL DEMINERALIZATION)	39
Bakradze A., Vadachkoria Z., Kvachadze I. ELECTROPHYSIOLOGICAL CORRELATES OF MASTICATORY MUSCLES IN NASAL AND ORONASAL BREATHING MODES	45
Borysenko A., Timokhina T., Kononova O. INDICATORS OF LOCAL IMMUNITY IN THE COMORBID COURSE OF CARIES AND GASTROESOPHAGEAL REFLUX DISEASE.....	48
Dolidze K., Margvelashvili V., Nikolaishvili M., Suladze T., Pkhaladze M. STUDY OF THE HYGIENIC CHARACTERISTICS OF THE ORAL CAVITY UNDER THE COMPLEX EFFECT OF PHOTODYNAMIC THERAPY AND TSKALTUBO SPRING WATER RADON HORMESIS.....	54
Танская О.А., Островский Ю.П., Курлянская Е.К., Валентюкевич А.В., Колядко М.Г. ОСНОВНЫЕ КРИТЕРИИ ОТБОРА ПАЦИЕНТОВ ПРИ ФОРМИРОВАНИИ ЛИСТА ОЖИДАНИЯ НА ТРАНСПЛАНТАЦИЮ СЕРДЦА	60
Yelshibayeva E., Dautov T., Rakhimzhanova R., Gutberlet M., Mardenkyzy D., Kozhakhmetova Zh., Saduakasova A. COMPUTED TOMOGRAPHY IN DETECTING FEATURES OF CORONARY ATHEROSCLEROSIS IN DIFFERENT ETHNIC GROUPS OF KAZAKHSTAN POPULATION.....	68
Podzolkov V., Safronova T., Nebieridze N., Loriya I., Cherepanov A. TRANSFORMING GROWTH FACTOR AND ARTERIAL STIFFNESS IN PATIENTS WITH UNCONTROLLED ARTERIAL HYPERTENSION	77
Gvasalia T., Kvachadze I., Giorgobiani T. SENSITIVITY TO MECHANICAL PAIN BASED ON SATIETY LEVELS IN WOMEN	83
Povoroznyuk V., Nishkumay O., Lazarieva K., Lazarev P. FEATURES OF BONE METABOLISM AND THEIR INFLUENCE ON ARTERIAL WALL STIFFNESS IN POSTMENOPAUSAL WOMEN WITH CONTROLLED UNCOMPLICATED HYPERTENSION	87
Solomonina N., Vacharadze K., Mgvdeladze G. CHARACTERISTICS OF DRUG RESISTANT TUBERCULOSIS IN GEORGIA (2015-2020).....	93

Abramidze T., Gotua M., Bochorishvili E., Melikidze N., Gamkrelidze A. CYPRESS POLLEN SENSITIZATION IN GEORGIA: CLINICAL AND MOLECULAR CHARACTERISTICS.....	101
Притыко Н.Г., Коваленко О.Е. ОСОБЕННОСТИ МОЗГОВОЙ ГЕМОДИНАМИКИ У ПАЦИЕНТОВ С СИНДРОМОМ ХРОНИЧЕСКОЙ ЦЕРЕБРАЛЬНОЙ ВЕНОЗНОЙ ДИСФУНКЦИИ И РАЗНЫМ УРОВНЕМ АРТЕРИАЛЬНОГО ДАВЛЕНИЯ.....	107
Chorna V., Makhniuk V., Pshuk N., Gumeniuk N., Shevchuk Yu., Khliestova S. BURNOUT IN MENTAL HEALTH PROFESSIONALS AND THE MEASURES TO PREVENT IT	113
Ratiani L., Gegechkory S., Machavariani K., Shotadze T., Sanikidze T., Intskirveli N. THE PECULIARITY OF COVID-19 GENOME AND THE CORONAVIRUS RNA TRANSLATION PROCESS AS A POTENTIAL TARGET FOR ETIOTROPIC MEDICATIONS WITH ADENINE AND OTHER NUCLEOTIDE ANALOGUES (REVIEW).....	119
Patarashvili L., Azmaipharashvili E., Jandieri K., Gvidiani S., Tsomaia K., Kikalishvili L., Sareli M., Chanukvadze I., Kordzaia D. LIVER EXTRACELLULAR MATRIX PECULIARITIES IN MAMMALS AND AVIANS.....	124
Tsomaia K., Azmaipharashvili E., Gvidiani S., Bebiashvili I., Gusev S., Kordzaia D. STRUCTURAL CHANGES IN RATS' LIVER DURING THE FIRST 2 WEEKS FOLLOWING 2/3 PARTIAL HEPATECTOMY	134
Gvianishvili T., Kakauridze N., Gogiashvili L., Tsagareli Z., Kurtanidze T. CORRELATION OF THYROID AUTOIMMUNITY WITH ATHEROSCLEROSIS EVALUATION IN HASHIMOTO'S THYROIDITIS.....	142
Kiknadze T., Tevdorashvili G., Muzashvili T., Gachechiladze M., Burkadze G. PHENOTYPIC CHARACTERISTICS OF RELAPSED LEIOMYOMA AND SMOOTH MUSCLE TUMORS OF UNCERTAIN MALIGNANCY POTENTIAL IN REPRODUCTIVE WOMEN.....	150
Pkhakadze G., Bokhua Z., Asatiani T., Muzashvili T., Burkadze G. STEM CELL INDEX IN THE PROGRESSION OF CERVICAL INTRAEPITHELIAL NEOPLASIA.....	157
Pidlisetsky A., Savosko S., Dolhopolov O., Makarenko O. PERIPHERAL NERVE LESIONS AFTER A MECHANICALLY INDUCED LIMB ISCHEMIA.....	165
Kolisnyk I., Voloshin O., Savchenko I., Yanchevskiy O., Rashidi B. ENZYMATIC ACTIVITY IN MICROSOMES, LIPID PEROXIDATION OF MICE HEPATOCYTES UNDER THE SODIUM FLUORIDE.....	169
Smagulova A., Katokhin A., Mambetpayeva B., Kulmaganbetova N., Kiyan V. A MULTIPLEX PCR ASSAY FOR THE DIFFERENTIAL DETECTION OF OPISTHORCHIS FELINEUS AND METORCHIS BILIS	176
Rigvava S., Karumidze N., Kusradze I., Dvalidze T., Tatrishvili N., Goderdzishvili M. BIOLOGICAL CHARACTERIZATION OF BACTERIOPHAGES AGAINST STREPTOCOCCUS AGALACTIAE	182
Deshko L., Udovenko Zh., Bulycheva N., Galagan V., Bulychev A. PROVISION OF THE RIGHT TO NON-INTERFERENCE WITH PRIVACY DURING MUSTER PROCESS WITH THE PARTICIPATION OF DOCTOR (FORENSIC EXPERT)	186
Теремецкий В.И., Николаенко Т.Н., Дидковская Г.В., Гмырин А.А., Шаповал Т.Б. КОНТРОЛЬ И НАДЗОР КАК СРЕДСТВА ПРЕДУПРЕЖДЕНИЯ И ВЫЯВЛЕНИЯ ПРАВОНАРУШЕНИЙ В СФЕРЕ ЗДРАВООХРАНЕНИЯ.....	192

THE PECULIARITY OF COVID-19 GENOME AND THE CORONAVIRUS RNA TRANSLATION PROCESS AS A POTENTIAL TARGET FOR ETIOTROPIC MEDICATIONS WITH ADENINE AND OTHER NUCLEOTIDE ANALOGUES (REVIEW)

¹Ratiani L., ¹Gegechkory S., ¹Machavariani K., ¹Shotadze T., ²Sanikidze T., ¹Intskirveli N.

¹Tbilisi State Medical University, The First University Clinic; ²Tbilisi State Medical University, Georgia

Coronaviruses (CoVs) belong to the family Coronaviridae. They are further subdivided into four genera: alphacoronavirus (α -CoV), betacoronavirus (β -CoV), gammacoronavirus (γ -CoV) and deltacoronavirus (δ -CoV) [6,48]. Both α - and β -CoVs infect mammals, while δ - and γ -CoVs infect birds. In early December 2019, the cases of pneumonia of unknown etiology were reported in Wuhan. After identifying the genome sequence of infected patients, it was revealed that the causative agent was a new type of coronavirus, namely SARS-CoV-2. Like SARS-CoV and MERS-CoV, the newly formed SARS-CoV-2 virus belongs to the group of β -CoVs.

The incubation period for SARS-CoV-2 is on average 3-7 days, although it can be up to 2-14 days [28, 43], which coincides with other known CoV incubation periods (e.g., SARS-CoV incubation period is on average 5 days, although it may increase to 2-14 days [4], the incubation period for MERS-CoV is approximately 5-7 days, although it can be up to 2-14 days [5, 34]. Asymptomatic patients can effectively transmit SARS-CoV-2 during the incubation period, [24].

The goal of the present article is to summarize and analyze the literature data concerning specific features of COVID-19 virus and to consider the potential targets for etiotropic therapy.

Genetic sequence. The genetic sequence of SARS-CoV-2 is 70% similar to the SARS-CoV sequence. Like the latter, SARS-CoV-2 uses the ACE2 (Angiotensin-Converting Enzyme/Enzyme 2) receptor to enter the cell and infect humans [46,54]. However, with the main antigen component, i.e. the S protein, it differs significantly from its predecessor. The S protein of SARS-CoV-2 binds to the human ACE2 receptor at 10- to 20-fold higher affinity, facilitating the spread of the virus among humans [44]. It should also be noted that the respiratory tracts are not the only route of transmission of the SARS-CoV-2 virus, and it is also transmitted even during close contact. Recent studies have shown that some patients with confirmed COVID-19 experience dyspeptic symptoms such as diarrhea, vomiting, nausea [22,32]. The enteric symptoms of COVID-19 are associated with the presence of the ACE2 receptors in the digestive tract [37].

SARS-CoV-2 uses the genomic RNA as a template to translate pp1a (polyprotein 1a) and pp1ab (polyprotein 1ab). These proteins (pp1a, pp1ab) produce nonstructural proteins (NSPs) in double-membrane vesicles (DMVs) to form the replication/transcription complex (RTC) [25].

Negative-stranded RNA (Coronavirus Genomic RNA (-)) is produced by replication of the complex. As a result of the transient transcription of its RNA-dependent RNA polymerase (RdRp), subgenomic RNAs of different lengths (sgRNAs) are produced [9]. Translation of each sgRNA results in the production of viral proteins, and replication of negative-stranded RNA (Coronavirus Genomic RNA (-)) yields positive-stranded RNA (+).

The SARS-CoV-2 genome and subgenome contain at least 6 open reading frames (ORFs). The first ORF (ORF1a/b) is 2/3 of the total genome length of SARS-CoV-2; it produces two polypeptides pp1a and pp1ab. Proteolytic cleavage of ORF1a/b results in producing 15/16 NSPs (nonstructural proteins), 4 structural proteins, and 5 complementary proteins (ORF3a, ORF6, ORF7a, ORF8, and ORF9) [23,47].

ORF1a encodes the production of pp1a, a molecular weight of which is 486 kDa. The pp1a protein contains P1pro (the papain-like protease), 3CLpro, and two membrane proteins MP1 (nsp4 - nonstructural protein 4) and MP2 (nsp6 - nonstructural protein 6).

NSP1 inhibits the synthesis of the cellular proteins in an infected cell. The cell is forced to regulate mainly viral protein synthesis. Moreover, the protein NSP1 does not allow the cellular antiviral proteins to aggregate that is necessary to stop the virus [17,40]. The function of the protein NSP2 has not been determined, only its ability to participate in the placement of endosomes along the cell has been identified [16,29]. The protein NSP3 performs two important functions: 1 - it provides the release of other viral proteins after which they begin to perform their own function; 2 - it changes the function of the proteins of the infected cell.

NSP3 is released by pp1a/1ab via a papain-like protease domain that is part of NSP3 itself [7]. NSP4, along with other proteins, is involved in the formation of fluid-containing blisters in an infected cell [38]. NSP5 is specialized in breaking down proteins, causing other NSPs to be activated and start to act [3,55,56]. NSP6 is involved in the formation of viral blisters along with the NSP3 and the NSP4 proteins [12,27]. NSP7 and NSP8 help NSP12 generate a copy of the virus RNA genome that gives rise to offspring viruses [14]. NSP9 penetrates into the nucleus with the help of small channels in the cell nucleus and influences the movement of molecules from the nucleus [13,51]. The protein NSP10, along with NSP16, disguises a viral gene and prevents the attack of antiviral proteins in human cells that have the ability to detect and destroy viral RNA [8,15]. The function of the NSP11 protein is unknown [2]. The protein NSP11, together with NSP12, concentrates nucleotides in the coronavirus genome. The ability of the antiviral drug remdesivir to interact with the coronavirus NSP12 protein has been identified; studies are being carried out regarding the widespread use of this drug in treatment [1,41].

In a normal state, the viral RNA is twisted. It is assumed that the NSP13 protein destroys viral RNA and thus makes it available for action on proteins involved in the production of new viral copies [11]. The NSP14 protein corrects the errors (incorrectly added nucleotides) made by the NSP12 protein during duplication of the coronavirus [15,52]. The protein NSP15 [53] supposedly protects the virus from the antiviral activities of the cell [19].

The underlying ORF1b is expressed as a pp1a fusion protein through a mechanism that involves the movement of the ribosomal backbone during translation [20,26]. The result is the protein pp1ab (\approx 790 kDa) that already contains ORF1b containing the helicase domain (nsp13) [39], exonuclease (nsp14), endoribonuclease (nsp15), and nsp16.

The remaining ORFs make up about one-third of the genome length, are located near the 3'-end, and encode at least four types of structural proteins:

- The E protein - a structural protein of the coronavirus membrane that forms the lipid vesicles of the virus. Inside the cell, these vesicles fix proteins that are involved in the human gene regulation process.

- The M protein - a membrane protein of coronavirus. It participates in the formation of the outer membrane of the virus;

- The S-S protein - forms the protective outer layer of the coronavirus RNA genome on the surface of the virus. In micrographs, the club-shaped spikes that stud the surface of coronaviruses are glycoproteins that give the appearance of a radiate crown. Their parts expand and attach to the ACE2 protein in human airway cells. Then it enters the cell.

- The N protein - protects viral RNA, promotes the internal stability of the virus. Most of the N proteins coalesce into a long helix and lead to the formation of the helical nucleocapsid.

So, the SARS-CoV (COVID-19) genome encodes the so-called “auxiliary proteins” that create a favorable environment inside the cell of the host organism, which promotes its multiplication. The ORF3a protein damages the host cell membrane, thus allowing new viruses to come out of the cell. This is what causes pneumonia – a symptom typical for COVID-19.

ORF6 inhibits the signals sent by the infected cell to the immune system, in addition, it inhibits the activity of some proteins in the cell.

When the virus starts coming out of an infected cell, the cell can bind it with the help of the tetherin protein. ORF7 is thought to reduce the supply of the tetherin protein in an infected cell, making it easier for viruses to leave the cell. It also provokes “cell suicide” (apoptosis) that significantly damages the lungs. The function of the ORF8 protein is unknown. ORF9b and ORF9c are coronavirus “auxiliary proteins”; ORF9b inhibits the action of the key protein interferon in the fight against cellular viruses; the function of the ORF9c protein is unknown.

Treatment. The general treatment strategy for COVID-19 involves bed rest and controlled intake of adjuvant medications. It is also recommended to maintain water and electrolyte balance while monitoring other vital parameters (heart rate, blood pressure, pulse, respiration rate, etc.). Some scientists are counting on the possible antiviral effects of IFN α .

Interferon-alpha (IFN α) belongs to the type I IFN family. It plays an important role in resistance to viral infections, inhibits viral infections by interfering with virus replication, and activates the host’s immune response. In vitro experiments

have shown that IFN α effectively inhibits SARS-CoV replication [46,47]. As revealed, IFN α protects cynomolgus macaques from SARS CoV [18,31,57]. Moreover, pilot clinical trials have shown positive therapeutic effects when using IFN α in patients with SARS [46].

Table lists the medications used to treat COVID-19 and shows the targets for their action.

As shown in Table, all of the antiviral drugs listed above have some antiviral (anti-SARS-CoV2) effects, and may have a certain result on the process of treating SARS-CoV2. The main targets of current medications are:

- Viral RNA-dependent RNA polymerase RdRp (Remdesivir inhibits RNA-dependent RNA polymerase (RdRp), thus blocking the production of viral proteins; However, in contrast, 3-5 exoribonucleases of the virus inhibit the action of remdesivir and reduce the antiviral effect of this drug);

- Viral 3CLpro or PLpro (the papain-like proteins lopinavir/ritonavir block already formed proteins, thus preventing further production of viral proteins);

- The Virus transmembrane S protein and transmembrane protease serine-2 [TMPRSS2] inhibitors (Arbidol and Camostat mesylate) can prevent the interaction of the S protein and the cellular receptor ACE2);

- The ACE 2 receptor on the host cell membrane that provides the entry point for the S protein (chloroquine and hydroxychloroquine inhibit endocytosis by increasing endosomal pH). Chloroquine can also inhibit RdRp by increasing intracellular zinc concentrations like remdesivir.

However, several key issues need to be emphasized: (1) The potential interaction of these antiviral drugs with other types of medications should be taken into account; (2) Side effects of two medication lopinavir/ritonavir should be considered (dyspepsia and liver damage); (3) Using three or more «antiviral» drugs with different mechanisms considering the side effects of some of them is controversial.

In addition to the medications listed above, research on the possibility of using antiviral antibodies in the plasma of recovered patients is being carried out intensively. Plasma therapy is commonly used in viral diseases such as influenza A (H5N1), poliomyelitis, and Ebola [10,50].

Table. List of the medications used to treat COVID-19 and the targets for their action

Therapeutic Target	Function	Potential Medications	References
RNA-dependent RNA polymerase (RdRp)	Coronavirus genome replication	Remdesivir and ribavirin. They have an ability to inhibit RdRp.	[36, 44]
The Papain-like protease PLpro	Converts viral polyprotein into a functional enzyme.	Lopinavir, protease inhibitor that can inhibit viral protease: 3CLpro/PLpro	[52]
The main protease 3CLpro	Converts viral polyprotein into a functional protein	Lopinavir	[21]
The S protein and TMPRSS2	The S protein of on the Virus surface that binds to the host surface ACE2 (angiotensin-converting enzyme/enzyme2) receptor. TMPRSS2 ‘supplements’ the S protein to bind to the ACE2 (angiotensin-converting enzyme/enzyme2) receptor.	Arbidol - It can prevent the interaction of the S protein and the ACE2 receptor and inhibit membrane fusion. Camostat mesylate inhibits TMPRSS2.	[33, 35]
ACE 2	A receptor on a host cell providing the entry point for the S-protein.	Chloroquine and hydroxychloroquine inhibit endocytosis by increasing endosomal pH.	[30,42]

Given the above-mentioned facts, in order to stop the spread of coronavirus infections and to avoid their damaging effects, it is promising to inhibit the production of NSPs of the viral origin, which is possible by replacing nucleotides, in particular adenine, in the viral RNA translation phase. The improved models of adenosine analogs such as remdesivir and NITD008 should be used for this purpose [49, 50] because both of them try to inhibit the viral replication process by inhibiting RdRp. In addition, it is known that 3'-5' exonucleases of SARS-CoV2 block the inhibitory effect of remdesivir on RdRp, promoting further replication of the virus. Therefore, it is necessary to create new modified/improved versions of remdesivir and other adenosine analogs.

Conclusion. The cases of SARS-CoV were first reported in 2002 and the virus quickly spread to 32 countries around the world. Ten years later, MERS-CoV became widespread in 2012, and eight years later, in 2020, a new viral infection SARS-CoV-2 emerged. It has been proven that SARS-CoV-2 enters the cell with the help of the ACE2 receptors. The fact that this type of receptors is found not only in the respiratory system but also in the liver tissues, the digestive system (small intestine, duodenum), testicles and kidneys, makes these organs highly vulnerable to SARS-CoV-2. The different group of drugs have been proposed in complex treatment of COVID-19. Despite of this coronavirus is still associated with high incidence of various complications and fatal outcome worldwide. According epidemiologic studies the most susceptible are the patients with accompanying diabetes, cardiovascular, respiratory system diseases and obesity. Potentially patients with intestinal microbiome disorders also may become vulnerable to COVID -19[32].

Given the global health threat caused by SARS-CoV-2, there is an urgent need for effective prevention and treatment of COVID-19 pneumonia, although finding drugs to treat pathogenic SARS-CoV-2 still remains a major problem. The medicines available to doctors around the world do not have a significant detrimental effect on the virus, as evidenced by the current epidemiological data. In initial stage the introduction of adenosine analog remdesivir against COVID-19 was considered as perspective drug[36]. This agent was approved or authorized in about 50 countries, including USA and EU, but currently there are controversial views regarding its ability to reduce mortality in COVID 19.

We suppose that if the improved versions of adenosine analogs (NITD008, Remdesivir, etc.) with more efficacy and safety are developed, the virus will not be able to have a detrimental effect on host cells because they (the improved versions of adenosine analogs) will have the ability to inhibit the virus translation process rather than RdRp. As a result, the virus will no longer be able to produce non-structural proteins (nsps) so important for the manifestation of viral activity.

REFERENCES

1. Adedeji AO, Lazarus H. (2016) Biochemical characterization of Middle East respiratory syndrome coronavirus helicase. *mSphere*. 7;1(5):e00235-16
2. Ahn DG, Choi JK, Taylor DR, Oh JW. Biochemical characterization of a recombinant SARS coronavirus nsp12 RNA-dependent RNA polymerase capable of copying viral RNA templates. *Arch Virol*. 2012; 157(11):2095-2104.
3. Angelini MM, Akhlaghpour M, Neuman BW, Buchmeier MJ. (2013) Severe acute respiratory syndrome coronavirus non-structural proteins 3, 4, and 6 induce double-membrane vesicles. *mBio*. 4(4):e00524-13:1-10.
4. Assiri A, Al-Tawfi JA, Al-Rabeeh AA, Al-Rabiah FA, et al. (2013) Epidemiological, demographic, and clinical characteristics of 47 cases of Middle East respiratory syndrome coronavirus disease from Saudi Arabia: a descriptive study. *Lancet Infect Dis*. 13: 752-61.
5. Backer JA, Klinkenberg D, Wallinga J. (2020) Incubation period of 2019 novel coronavirus (2019-nCoV) infections among travellers from Wuhan, China, 20-28 January. *Euro Surveill*. 25(5): 1-6.
6. Banerjee A, Kulcsar K, Misra V, Frieman M, Mossman K. (2019) Bats and Coronaviruses. *Viruses*. 1(1). 41:1-15.
7. Beachboard DC, Anderson Daniels JM, Denison MR. (2015) Mutations across murine hepatitis virus nsp4 alter virus fitness and membrane modifications. *J Virol*. 89(4):2080-2089.
8. Bouvet M, Imbert I, Subissi L, Gluais L, Canard B, Decroly E (2012). RNA 3' end mismatch excision by the severe acute respiratory syndrome coronavirus nonstructural protein nsp10/nsp14 exonuclease complex. *Proc Natl Acad Sci USA*;109(24): 9372-9377.
9. Chan JF-W, Kok K-H, Zhu Z, Chu H, et al. (2020) Genomic characterization of the 2019 novel human-pathogenic coronavirus isolated from a patient with atypical pneumonia after visiting Wuhan. *Emerg Microbes Infect*. 9:221-236.
10. Charles R Rinaldo Jr. (2005) Passive immunization against poliomyelitis: the Hammon gamma globulin field trials, 1951-1953. *Am J Public Health*;95(5):790-9.
11. Chen Y, Cai H, Pan J, Xiang N, Tien P, Ahola T, Guo D. (2009) Functional screen reveals SARS coronavirus nonstructural protein nsp14 as a novel cap N7 methyltransferase. *Proc Natl Acad Sci USA*. 3;106(9):3484-3489.
12. Cottam EM, Whelband MC, Wileman T. (2014) Coronavirus NSP6 restricts autophagosome expansion. *Autophagy*. 10(8):1426-1441
13. Decroly E, Debarnot C, Ferron F, et al. (2011) Crystal structure and functional analysis of the SARS coronavirus RNA cap 2'-O-methyltransferase nsp10/nsp16 complex. *PLOS Pathog*. 7(5):e1002059:1-14.
14. Egloff MP, Ferron F, Campanacci V, et al. (2004) The severe acute respiratory syndrome-coronavirus replicative protein nsp9 is a single-stranded RNA-binding subunit unique in the RNA virus world. *Proc Natl Acad Sci U S A*;101(11):3792-3796.
15. Fang SG, Shen H, Wang J, Tay FPL, Liu DX. (2008) Proteolytic processing of polyproteins 1a and 1ab between non-structural proteins 10 and 11/12 of coronavirus infectious bronchitis virus is dispensable for viral replication in cultured cells. *Virology*. 379(2):175-180.
16. Gadlage MJ, Graham RL, Denison MR. (2008) Murine Coronaviruses Encoding nsp2 at Different Genomic Loci Have Altered Replication, Protein Expression, and Localization. *Journal Virology*; 92(23):11964-11969.
17. Graham RL, Sims AC, Brockway SM, Baric RS, Denison MR. (2005) The nsp2 replicase proteins of murine hepatitis virus and severe acute respiratory syndrome coronavirus are dispensable for viral replication. *Journal of Virology*. 79(21):13399-411.
18. Haagmans BL1, Kuiken T, Martina BE, Fouchier RA, Rimmelzwaan GF, van Amerongen G, van Riel D, de Jong T, Itamura S, Chan KH, Tashiro M, (2004) Pegylated interferon- α protects type 1 pneumocytes against SARS coronavirus infection in macaques. *Nature Medicine*, 10(3):290-293.
19. Herold J, Raabe T, Schelle-Prinz B, Siddell SG (1993) Nucleotide Sequence of the Human Coronavirus 229E RNA Polymerase Locus. *Virology*. 195:680-691.
20. Herold J, Siddell SG. (1993) An 'elaborated' pseudoknot

is required for high frequency frameshifting during translation of HCV 229E polymerase mRNA. *Nucleic Acids Res.* 21(25): 5838–5842.

21. Hoffmann M, Kleine-Weber H, Schroeder S, Krüger N, et al. (2020) SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor *Cell.* 181(2):271-280.e8.

22. Holshue ML, DeBolt C, Lindquist S, Lofly KH, Wiesman J, Bruce H. (2020) First case of 2019 novel coronavirus in the United States. *N Engl J Med.*; 382(10):929–936.

23. Huang C, Lokugamage KG, Rozovics JM, Narayanan K, Semler BL. (2011) SARS Coronavirus nsp1 Protein Induces Template-Dependent Endonucleolytic Cleavage of mRNAs: Viral mRNAs Are Resistant to nsp1-Induced RNA Cleavage. *PLoS Pathog.* 8(11):e1002433:1-18.

24. Hui DS. (2020) The continuing 2019-nCoV epidemic threat of novel coronaviruses to global health — The latest 2019 novel coronavirus outbreak in Wuhan, China. *Int J Infect Dis.* 91:264–266.

25. Hussain S, Pan J, Chen Y, Yang Y, Xu J, Peng Y, et al. (2005) Identification of Novel Subgenomic RNAs and Noncanonical Transcription Initiation Signals of Severe Acute Respiratory Syndrome Coronavirus. *J Virol.* 79(9):5288–5295.

26. Ivanov K.A., Thiel V, Dobbe JC, van der Meer Y, Sijder EJ, Ziebuhr J. (2004) Multiple enzymatic activities associated with severe acute respiratory syndrome coronavirus helicase. *J Virol.*;78(11):5619-32.

27. Kirchdoerfer RN, Ward AB. (2019) Structure of the SARS-CoV nsp12 polymerase bound to nsp7 and nsp8 cofactors. *Nat Commun.* 28;10(1):2342.

28. Lauer SA, Grantz KH, Bi Q, Jones FK, Zheng Q, et al. (2020) The Incubation Period of Coronavirus Disease 2019 (COVID-19) From Publicly Reported Confirmed Cases: Estimation and Application. *Ann Intern Med.*; 5;172(9): 577-582.

29. Lei J, Kusov Y, Hilgenfeld R. (2018) Nsp3 of coronaviruses: structures and functions of a large multi-domain protein. *Antiviral Res.* 140:58–74.

30. Liu J, Cao R, Xu M, Wang X, Zhang H, Hu H, et al. (2020) Hydroxychloroquine, a less toxic derivative of chloroquine, is effective in inhibiting SARS-CoV-2 infection in vitro. *Cell Discovery.* 6:16: 1-4.

31. Loutfy MR, Blatt LM, Siminovitch KA, Ward S, Wolff B, Lho H, et al. (2003) Interferon alfacon-1 plus corticosteroids in severe acute respiratory syndrome: a preliminary study *JAMA.* 24;290(24):3222-8.

32. Ma C, Cong Y, Zhang H. (2020) COVID-19 and the Digestive System *Am J Gastroenterol.* ;115(7):1003-1006.

33. Rameshwar U. Kadam and Ian A. Wilson. Structural basis of influenza virus fusion inhibition by the antiviral drug Arbidol. *PNAS.* 2017; 114 (2): 206-214.

34. Rothe C, Schunk M, Sothmann P, Bretzel G, Froeschl G, et al. (2020) Transmission of 2019-nCoV Infection from an Asymptomatic Contact in Germany. *N Engl J Med.* 392.10:970–971.

35. Sandro GVR, Wilson CS. (2020) Clinical trials on drug repositioning for COVID-19 treatment *Rev Panam Salud Publica.* 40:1-6

36. Sheahan TP, Sims AC, Leist SR, Schäfer A, Won J, Brown AJ, Montgomery SA, Hogg A, Babusis D, Clarke MO, et al. (2020) Comparative therapeutic efficacy of remdesivir and combination lopinavir, ritonavir, and interferon beta against MERS-CoV. *Nat. Commun.* 10;11(11):222-224.

37. Snijder EJ, van der Meer Y, Zevenhoven-Dobbe J, Onderwater JJM, et al. (2006) Ultrastructure and Origin of Membrane Vesicles Associated with the Severe Acute Respiratory Syndrome

Coronavirus Replication Complex. *J Virol.*; 80(12):5927–5940.
38. Stobart CC, Sexton NR., Munjal H, Lu X, Molland KL, et al. (2013) Chimeric exchange of coronavirus nsp5 proteases (3CL-pro) identifies common and divergent regulatory determinants of protease activity. *Journal of Virology.* 87(23):12611–12618.

39. Ströher U, DiCaro A, Li Y, Strong JE, Aoki F, Plummer F. (2004) Severe acute respiratory syndrome-related coronavirus is inhibited by interferon-alpha. *J Infect Dis.*; 189:1164-1167.

40. Tanaka T, Kamitani W, DeDiego ML, Enjuanes L, Matsuura Y. (2012) Severe acute respiratory syndrome coronavirus nsp1 facilitates efficient propagation in cells through a specific translational shutoff of host mRNA. *J Virol.* 86(20):11128–11137.

41. te Velthuis AJW, Arnold JJ, Cameron CE, van den Worm-SHE, et al. The RNA polymerase activity of SARS-coronavirus nsp12 is primer dependent *Nucleic Acids Res.* 2010;38(1):203-14.

42. van Griensven J, Edwards T, de Lamballerie X, Semple MG, Gallian P, et al. (2016) Evaluation of Convalescent Plasma for Ebola Virus Disease in Guinea. *N Engl J Med.* 374(1):33–42

43. Varia M, Wilson S, Sarwal S, McGeer A, et al. (2003) Investigation of a nosocomial outbreak of severe acute respiratory syndrome (SARS) in Toronto, Canada. *CMA.* 169(4): 285–292.

44. Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, Wang B, Xiang H, et al. (2020) Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus–Infected Pneumonia in Wuhan, China. *JAMA.*; 323(11):1061–1069.

45. Wang M, Cao R, Zhang L, Yang X, Liu J, Xu M. (2020) Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res.*30: 269–271.

46. Wrapp D, Wang N, Corbett KS, Goldsmith JA, Hsieh CL, Abiona O. (2020) Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. *Science.*367:1260–1263.

47. Wu A, Peng Y, Huang B, Ding X, Wang X, et al. (2020) Genome Composition and Divergence of the Novel Coronavirus (2019-nCoV) Originating in China. *Cell Host Microbe.*; 27(3):325-328.

48. Yang D, Leibowitz JL (2015) The structure and functions of coronavirus genomic 3' and 5' ends. *Virus Res.* 206: 120–133.

49. Yina, Z, Chena Y-L, Schula W, Wanga Q-Y, Gua F, et al. (2009) An adenosine nucleoside inhibitor of dengue virus. *PNAS.* 106(48):20435–20439

50. Yong-Qiang D., Zhang N-N, Li C-F, Tian M, et al. (2016) Adenosine Analog NITD008 Is a Potent Inhibitor of Zika Virus. *Open Forum Infect Dis.* 3(4):1-4.

51. Zeng Z, Deng F, Shi K, et al. (2018) Dimerization of Coronavirus nsp9 with Diverse Modes Enhances Its Nucleic Acid Binding Affinity. *J Virol.*; 16;92(17):e00692-18

52. Zhang L, Lin D, et al. (2020) α -Ketoamides as Broad-Spectrum Inhibitors of Coronavirus and Enterovirus Replication: Structure-Based Design, Synthesis, and Activity Assessment. *Journal of Medicinal Chemistry.* 63(9):1-14.

53. Zhang L, Li L, Yan L, Ming Z, Jia Z, Lou Z, Rao Z (2018) Structural and biochemical characterization of endoribonuclease Nsp15 encoded by middle east respiratory syndrome coronavirus. *J Virol.*; 92:e00893–18.

54. Zhou P, Yang XL, Wang XG, Hu B, Zhang L, Zhang W. (2020) A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature.* 579:270–273.

55. Zhu X, Fang L, Wang D, et al. (2017) Porcine deltacoronavirus nsp5 inhibits interferon- β production through the cleavage of NEMO. *Virology.* 502:33–38.

56. Zhu X, Wang D, Zhou J, et al. (2017) Porcine Deltacoronavirus nsp5 Antagonizes Type I Interferon Signaling by Cleaving STAT2. *J Virol.* 91(10):e00003-17.

57. Zorzitto J, Galligan CL, Ueng JJ, Fish EN. (2006) Characterization of the antiviral effects of interferon- α against a SARS-like coronavirus infection in vitro. *Cell Res.* 16:220-229.

SUMMARY

THE PECULIARITY OF COVID-19 GENOME AND THE CORONAVIRUS RNA TRANSLATION PROCESS AS A POTENTIAL TARGET FOR ETIOTROPIC MEDICATIONS WITH ADENINE AND OTHER NUCLEOTIDE ANALOGUES (REVIEW)

¹Ratiani L., ¹Gegechkory S., ¹Machavariani K., ¹Shotadze T., ²Sanikidze T., ¹Intskirveli N.

¹Tbilisi State Medical University, The First University Clinic; ²Tbilisi State Medical University, Georgia

Despite the multifaceted effects of the medicines provided for COVID-19 treatment, the number of the infected and mortality of patients increases which demonstrates the insufficient effectiveness of drugs used to fight coronavirus infections in medical practice, and clearly shows the need to develop new treatment tactics. In this review article are summarized and analyzed the literature data concerning specific features of COVID 19. Particular attention is given to genetic characteristic of this virus, to mechanism of its invasion into the human organism, replication and interaction with ACE-2 receptors, as well as to the basic targets for the action of existing drugs with antiviral activity against COVID-19.

Currently, the following medications are used to treat COVID-19: remdesivir, chloroquine, hydroxychloroquine (HCQ), ribavirin, lopinavir/ritonavir. According to a recent theory of coronavirus treatment, the starting point for the

mechanism of action of a potential etiotropic drug is the inhibition of the coronavirus main protease (Mpro/3CLpro) and the papain-like protease (PLpro). Among the drugs listed above, lopinavir acts through this mechanism but is characterized by severe side effects. It is emphasized that remdesivir as adenosine analog provides inhibitory action on RNA dependent RNA-Polymerase, but there are controversial views about reduction in mortality during using of this drug against COVID-19.

The present paper discusses the mechanism of action of a potential etiotropic drug against coronavirus, which implies the replacement of the nucleotides involved in the process of translation of the virus with their analogs with the aim to "inhibit" the ribosome and block the production of viral proteins.

Keywords: COVID-19, Genetic sequence, etiotropic drug, ribosome.

РЕЗЮМЕ

ОСОБЕННОСТИ ГЕНОМА COVID-19 И ТРАНСЛЯЦИОННЫЙ ПРОЦЕСС РНК КОРОНАВИРУСА КАК ПОТЕНЦИАЛЬНАЯ МИШЕНЬ ДЛЯ ЭТИОТРОПНОЙ ТЕРАПИИ АДЕНИНОМ И РАЗНЫМИ АНАЛОГАМИ НУКЛЕОТИДОВ (ОБЗОР)

¹Ратиани Л.Р., ¹Гегечкори С.С., ¹Мачавариани К.Ш., ¹Шотадзе Т.Г., ²Саникидзе Т.В., ¹Инцкирвели Н.А.

¹Тбилисский государственный медицинский университет, Первая университетская клиника;
²Тбилисский государственный медицинский университет, Грузия

Несмотря на многочисленные эффекты лекарственных средств, применяемых для лечения COVID-19, количество инфицированных и смертность пациентов увеличивается, что свидетельствует о недостаточной эффективности препаратов, применяемых в медицинской практике для борьбы с коронавирусными инфекциями, и необходимости разработки новой тактики лечения.

В настоящей обзорной статье суммированы и проанализированы данные литературы, касающиеся специфических черт коронавируса. Особое внимание уделяется генетической характеристике этого вируса, механизму его инвазии в человеческий организм, репликации и взаимодействию с АКФ-2 рецепторами, также как и основным мишеням для действия существующих лекарств, обладающих противовирусной активностью против коронавируса.

В настоящее время для лечения COVID-19 используются следующие препараты: ремдесивир, хлорохин, гидроксихлорохин (HCQ), рибавирин, лопинавир/ритонавир. Согласно существующей теории лечения коронавируса, отправной точкой для механизма действия потенциального этиотропного препарата является ингибирование основной протеазы коронавируса (Mpro/3CLpro) и папаин-подобной протеазы (PLpro). Среди вышеперечисленных препаратов лопинавир действует посредством этого механизма, однако характеризуется серьезными побочными эффектами. В данной статье обсуждается механизм действия потенциального этиотропного препарата против коронавируса, который подразумевает замену нуклеотидов, участвующих в процессе трансляции вируса, их аналогами с целью «ингибировать» рибосомы и блокировать производство вирусных белков.

რეზიუმე

კოვიდ-19-ის გენომის თავისებურებანი და კორონავირუსის რნმ-ის ტრანსლაციური პროცესი, როგორც პოტენციური სამიზნე ადენინით და ნუკლეოტიდების სხვადასხვა ანალოგებით ეტიოტროპული თერაპიისთვის (მიმოხილვა)

¹ლ.რატიანი, ¹ს.გეგუკორი, ¹კ.მაჭავარიანი, ¹თ.შოთაძე, ²თ.სანიკიძე, ¹ნ.ინწკირველი

¹თბილისის სახელმწიფო სამედიცინო უნივერსიტეტი, პირველი საუნივერსიტეტო კლინიკა;
²თბილისის სახელმწიფო სამედიცინო უნივერსიტეტი, საქართველო

COVID-19-ის სამკურნალოდ მოწოდებული მედიკამენტების მრავალმხრივი ეფექტების მიუხედავად, ინფიცირებულთა რიცხვი და პაციენტთა სიკვდილიანობა მატულობს, რაც ცხადყოფს კორონავირუსის ინფექციებთან ბრძოლის სამკურნალოდ გამოყენებული მედიკამენტების არასაკმარის ეფექტურობას და მკურნალობის ახალი ტაქტიკის შემუშავების აუცილებლობას. სტატიაში სუმირებული და გაანალიზებულია ლიტერატურული მონაცემები, რომლებიც ეხება კორონავირუსის სპეციფიკურ ნიშნებს, კერძოდ, განსაკუთრებული ყურადღება გამახვილებულია ამ ვირუსის გენეტიკურ მახასიათებლებზე, ადამიანის ორგანიზმში მისი ინვაზიის, რეპლიკაციის მექანიზმზე და ამფ-2 რეცეპტორებთან ურთიერთქმედებაზე, ისევე როგორც კორონავირუსის საწინააღმდეგოდ მოქმედი ანტივირუსული ეფექტის მქონე არსებული პრეპარატების ძირითად სამიზნეებზე.

ამჟამად COVID-19-ის სამკურნალოდ გამოიყენება

შემდეგი მედიკამენტები: რემდესვირი, ქლოროქინი, ჰიდროქსიქლოროქინი (HCQ), რიბავირინი, ლოპინავირი/რიტონავირი. კორონავირუსის მკურნალობის ბოლოდროინდელი თეორიის თანახმად, პოტენციური ეტიოტროპული პრეპარატის მოქმედების მექანიზმის ამოსავალი წერტილი არის კორონავირუსის მთავარი პროტეაზას (Mpro/3CLpro) და პაპაინის მსგავსი პროტეაზას (PLpro) დათრგუნვა. ზემოთ ჩამოთვლილ მედიკამენტებს შორის ლოპინავირი მოქმედებს ამ მექანიზმის საშუალებით, მაგრამ მას ახასიათებს მწვავე გვერდითი მოვლენები.

წინამდებარე ნაშრომში განხილულია კორონავირუსის საწინააღმდეგოდ პოტენციური ეტიოტროპული პრეპარატის მოქმედების მექანიზმი, რაც გულისხმობს ვირუსის რეპლიკაციის პროცესში ჩართული ნუკლეოტიდების ჩანაცვლებას მათი ანალოგებით რიბოსომის “დათრგუნვის” და ვირუსული ცილების წარმოების ბლოკირების მიზნით.

LIVER EXTRACELLULAR MATRIX PECULIARITIES IN MAMMALS AND AVIANS

¹Patarashvili L., ^{1,4}Azmaipharashvili E., ³Jandieri K., ¹Gvidiani S., ^{1,2}Tsomaia K.,
³Kikalishvili L., ⁵Sareli M., ³Chanukvadze I., ^{1,2}Kordzaia D.

¹Ivane Javakhishvili Tbilisi State University (TSU), Faculty of Medicine; ²Alexandre Natishvili Institute of Morphology, TSU;

³Tbilisi State Medical University; ⁴Institute of Clinical Oncology, Tbilisi, Georgia;

⁵Chaim Sheba Medical Center at HaShomer, Department of Surgical Oncology (Surgery C), Tel-Aviv, Israel

The extracellular matrix - the connective tissue framework of the liver - on the one hand, determines the shape of the organ, and on the other hand, creates specialized compartments for blood and lymphatic vessels and nerves, as well as cell populations, the synergy of which determines the various functioning of the organ. The liver is the largest and heaviest parenchymal organ, and an appropriate matrix design is required to maintain its shape and fix it on the abdominal walls [1]. The liver has a dual blood supply (arterial and portal), and the connective tissue spaces containing these vessels are built with this factor in mind. Unlike other organs, in which there are three circulating fluids and, therefore, there are three compartments for the microcirculation, four fluids circulate in the liver: blood, bile, interstitial juice, and lymph [2]. At the same time, the liver produces more lymph than any other organ (up to 50% of the total amount of lymph in the body). Thus, the liver matrix forms a highly complex but strongly regulated labyrinth in which liver cells, blood

vessels, bile ducts, lymphatic ducts, and tissue fluid have their own but closely interconnected compartments [3-5].

The study of the liver connective tissue skeleton dates back to the 17th century. Pursuant to Couinaud [6], in 1640 Walaeus described the connective tissue sheath, which wraps the portal vein, the hepatic artery, the bile duct, the lymphatic duct, and the nerves entering and leaving the liver connects to the capsule of the liver and hepatoduodenal ligament. Walaeus sheath originates from the vasculobiliary sheath (Glisson's capsule) and is not derived from the peritoneum or the capsule of the liver (Laennec's capsule). Besides, the separation between Laennec's capsule and the Walaeus sheath can be seen microscopically at the hepatic hilum [6], where the Walaeus sheath forms a thick plate at the inferior part of the liver referred to as the hilar (portal) plate [7].

The portal pedicle wrapped by the Walaeus sheath continues inside the organ, as the so-called Glissonian Pedicals [8].